

Movement response of Orange-Vaal largemouth yellowfish (*Labeobarbus kimberleyensis*) to water quality and habitat features in the Vaal River, South Africa

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Abstract Many threatened fish species that utilize riverine habitats are faced with habitat degradation and subsequent deterioration in their ecological surroundings. Habitat degradation is a consequence of water quality parameters associated with anthropogenic activities including mining, industrial, agricultural and urban activities. We examined how the movement behaviour of radio-tracked Orange-Vaal largemouth yellowfish (*Labeobarbus kimberleyensis*) responded to a suite of water quality chemical parameters and habitat features in the Vaal River, South Africa. We found that the probability of their movement increased with a decrease in water clarity, presence of emergent and overhanging marginal vegetation and fast flowing rapids. High mobility in conditions of low water clarity was probably related to low prey capture success of this piscivorous fish. High movement of largemouth yellowfish in emergent and overhanging marginal vegetation areas and rapid habitat biotopes were attributed to accessibility of prey within these important cover features. When water quality parameters were considered, the probability of largemouth yellowfish movement increased with increasing levels of dissolved chloride (Cl) and silicon

(Si), whereas movement decreased with high phosphate (PO_4) concentrations and increased with alkalinity levels in the river. High nutrient levels associated with eutrophication caused reductions in the movement of largemouth yellowfish. The association of increased movement of largemouth yellowfish with increasing Cl and Si is indicative of degraded habitat condition in the Vaal River system. Our study showed the importance of monitoring fish behavioural movement patterns to multiple environmental parameters, as these fish are important ecological indicators when appropriate conservation and management plans of freshwater ecosystems are required.

Keywords Ecosystem indicator · Freshwater ecosystem · Radio telemetry · Environmental parameters · Vaal River · South Africa

Introduction

An important factor of environmental monitoring is determining how water quality, physical habitat and other anthropogenic disturbances affect the behaviour and physiology of organisms (Aparicio and Sostoa 1999; Harvey et al. 1999; Gilliam and Fraser 2001; Harrison and Whitfield 2004). Fish are considered as one of the best ecological indicators of freshwater ecosystem health as they exhibit distinct behavioural, physiological and morphological responses to environmental stressors (Cooke and Cowx 2004; Harrison and Whitfield 2004, 2006; Elliott et al. 2007). Monitoring

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the movement patterns of aquatic organisms, such as fish, can offer an intuitive understanding of their distribution as well as tolerance to associated environmental conditions (Fausch et al. 2002; Gowan and Fausch 2002; Albanese et al. 2004; Petty and Grossman 2004; Gowan 2007; Roberts and Angermeier 2007). Thus, this knowledge could improve our understanding of river fish ecology amidst increasing anthropogenic impacts worldwide (Gowan et al. 1994; Roberts and Angermeier 2007).

Locomotion of fish is a dynamic biological process indicating the general health and physiological response of fish (Schreck et al. 1997; Cooke et al. 2003). In fish species, chronic exposure to multiple stressors reduces swimming ability, growth and habitat condition which may impair fish health and possibly result in mortality (Bervoets and Blust 2003; Wepener et al. 2011). In aquatic ecosystems the evaluation of excessive movement of species beyond normal ranges is challenging but studies have shown that it can be determined using movement rate (Gerhardt 2007). Excessive anthropogenic effects reduce fitness and survival potential in many species (Brind'Amour and Lobry 2009). Fish health is influenced by poor water quality, accumulation of heavy metals/toxicants and alteration in habitat structure from polluted environments (Gerhardt 2007). The movement responses of specialist species to environmental pollutants can affect physiology, produce physical abnormalities and lead to mortalities (Gerhardt 2007; Brind'Amour and Lobry 2009). The first response to excessively stressful environments may include avoidance of non-optimal conditions, followed by physiological stress where respiratory, metabolic or excretory rates may increase (Dallas and Day 2004). Telemetry techniques allow fish tracking in response to environmental stressors (Lucas et al. 1993; Cooke et al. 2003; O'Brien et al. 2012a, 2013a, b). Considering the importance of movement processes, studies have linked movement of freshwater river fish to ecological factors such as physical features, habitat structure and chemical variables (Gowan et al. 1994; Gilliam and Fraser 2001; Gerhardt 2007).

Multiple physico-chemical variables including water clarity, depth, pH, salinity, conductivity, nutrients, organic enrichment, trace metals and other toxicants, flow and habitat structure potentially influence the wellbeing of aquatic ecosystems (Dallas and Day 2004; Villiers

and Thiar 2007; Monette and McCormick 2008; Wepener et al. 2011). In South Africa, a large proportion of river basins (60%) have been exposed to habitat destruction (Revenge et al. 2005), affecting their primary functions and services including nutrient recycling, waste purification and maintaining large biodiversity (Palmer et al. 2005; Revenge et al. 2005). In particular, the Vaal River is located in the main mining and industrial areas of South Africa (Wepener et al. 2011). In the past, the water quality in this river has decreased significantly due to mixing of run-off waste such as salt load, nutrients and agriculture pesticides from the metropolitan area leading to mass mortality, particularly of yellowfish species (*Labeobarbus* spp.) which are habitat specialists in the Vaal River system (McCarthy et al. 2007; Villiers and Thiar 2007; McCarthy and Pretorius 2009; Wepener et al. 2011). The health of aquatic ecosystems can be studied using biological indicators such as socially and economically important apex predators, like the Orange-Vaal largemouth yellowfish (*Labeobarbus kimberleyensis*) (hereafter referred to as largemouth yellowfish), as they are sensitive to environmental changes (O'Brien et al. 2011, 2013a, b).

The largemouth yellowfish is an endemic and a high priority conservation species in South Africa region (Impson et al. 2008). It is one of the largest cyprinid fish in southern Africa. Moreover, the IUCN has categorized this species as "Near Threatened" (Impson and Swartz 2007) primarily due to reduction in its habitats and population size in its distribution range, and its sensitivity to flow modifications and poor water quality (Impson and Swartz 2007; Skelton and Bills 2007; De Villiers and Ellender 2008; Impson et al. 2008; DWA 2010; Ellender et al. 2012). In particular, the rapid biotope habitat (rapidly changing water flow) can also change the movements of fish as they find inaccessible habitats or find refuge from high current velocities (Albanese et al. 2004; O'Brien et al. 2013a, b). Previous studies have mainly focused on largemouth yellowfish habitat utilization (O'Brien et al. 2011, 2013a, b) but none on evaluating their movement response to water quality and habitat change. To fill this knowledge gap, we aimed to understand the movement behaviour of this species according to a suite of water quality and habitat features. We hypothesised that movement behaviour largemouth yellowfish in the Vaal River, South Africa, is influenced by a suite of water quality chemical

parameters and habitat features, though in different ways. Based on this we had the following predictions:

- that high yellowfish activity/movement occurs where they are more likely to find prey species within a range of habitats such as emergent and overhanging marginal vegetation in the littoral zone and bank of the river where they can presumably meet their resource needs.
- that yellowfish movement rates would increase as water clarity declined as this may reduce the feeding ability of piscivores such as yellowfish.
- that yellowfish movement rate may increase within the rapid biotope which allow them to exploit previously inaccessible habitats or find refuge from high current velocities.
- that yellowfish will increase their movement to increasing water conductivity while increased nutrient loading can lead to elevated ammonia and nitrite concentration which are toxic to many aquatic organisms and such nutrient enrichment can lead to a rapid numerical increase in fast-growing plant, thus reducing fish movement (Herbert and Steffensen 2005; Zhang et al. 2012).

Materials and methods

Study site

The Vaal River (upstream, 26°54'0.99"S; 27°26'45.39" E; downstream, 27°41'55.93"S; 26° 5'22.31"E, Fig. 1) is one of Africa's most important rivers providing water supply to Gauteng Province, the economic heartland of South Africa, supporting 42% of the urban population in the region (Braun and Rogers 1987). The river rises on the Drakensberg escarpment and flows ~900 km to the confluences with the Orange River (Braun and Rogers 1987; Bertasso 2004, Fig. 1). In South Africa, the catchments of the Vaal River spans approximately 192,000 km² and it is exposed to discharges from gold and coal mines, industry and sewage-treatment plants (DWA 2010; Wepener et al. 2011). Also, construction of weirs, dams/ impoundments and small manmade lakes has altered the natural water flow (Wepener et al. 2011). Our study was done in a reach of the middle Vaal River, southwest of Johannesburg (Fig. 1), on a 190 km section from Parys to upstream of Bothaville (Fig. 1). The study area is characterised by a sequence of moderately deep

<3 m mud and sand dominated pools with intermittent shallow >1 m cobble, boulder and bedrock dominated rapid habitats. The slight gradient of the reach ranges between 0.2% to 0.6% which results in velocities between 0 and 1.5 m/s. The study area is under high anthropogenic pressures which have local impacts on water quality, habitat quality, and river flow (DWA 2010; Wepener et al. 2011; O'Brien et al. 2013a). For the study area details, see O'Brien et al. (2013a, b). The area receives summer rainfall and mean annual rainfall of ~500 mm to 600 mm.

Study species

In this riverine system, the largemouth yellowfish is considered an apex predator that reaches over 20 kg. It primarily occurs in deep pools of large rivers and also in slow-moving water upstream sections of weirs and structures (De Villiers and Ellender 2007; Ellender et al. 2012). This apex predator, initially feeds on insects and small crustaceans as a juvenile, then when larger in size it is a more carnivorous and opportunistic hunter (O'Brien and De Villiers 2011).

Data collection

In total, 27 adult largemouth yellowfish were captured using gill netting, fyke trapping and electrofishing (boat-mounted electrofisher (1 kV) techniques (O'Brien et al. 2013a, b). Fishing effort for gill nets (93 mm mesh, 25 m length nets) was two nights for 12 h, five 12 h night efforts deployed in the evening and early morning for large fyke net traps (22 mm mesh with two 35 m wings), and purse dragged eight times for a large seine net (22 mm mesh, 35 m length). Every fish was tagged with radio transmitters (Weight 20 g; Advanced Telemetry Systems Inc. (ATS), Isanti, MN, USA) resulting in a tag burden of 2% of their body mass. Each captured largemouth yellowfish was anaesthetised by placement in a 50 L water filled container with either 2-phenoxy-ethanol (0.4 mL/L) or clove oil (0.1 mL/L) added. We attached transmitters dorso-laterally through the musculature just below the dorsal fin using stainless steel wire [see detailed tagging procedure in O'Brien et al. (2013a, b)]. We injected an antibiotic (Terramycin® containing oxy-tetracycline) (1 ml/kg) and applied Betadine (Mundipharma, RSA) on touched areas of the fish and wound-care gel (Aqua Vet) on wounds to treat and minimize their risk of

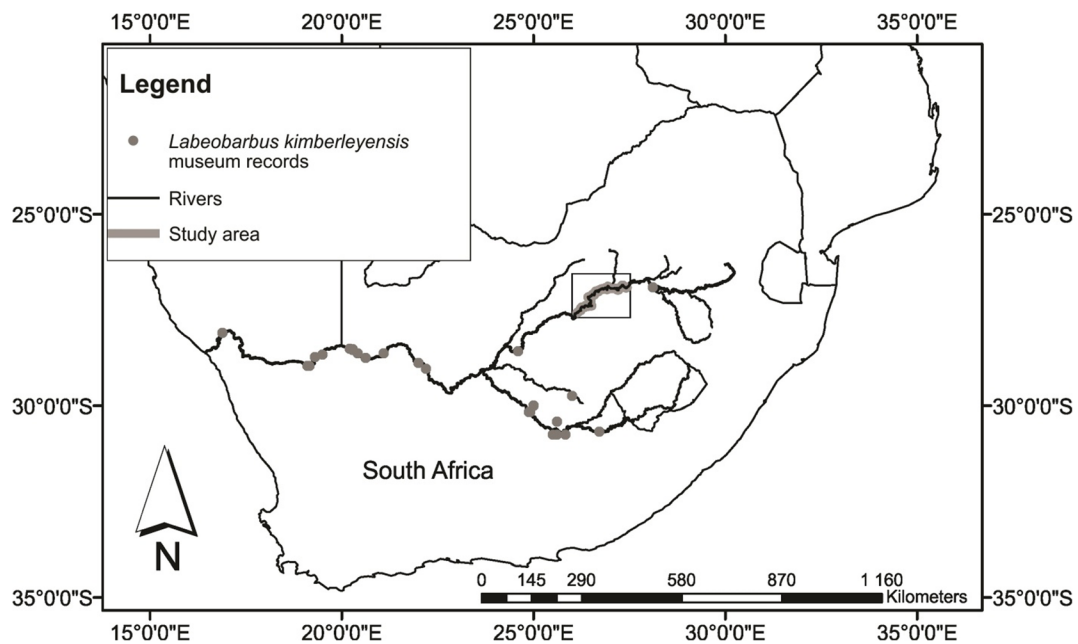


Fig. 1 Map of the survey regions showing the distributions of largemouth yellowfish *Labeobarbus kimberleyensis* (LKIM) on the Vaal River, southern Africa

infections. The total body length, fork length, girth length (mm) and body weight (g) were recorded from each tagged yellowfish and it was placed in circulating water within a 50 L water container until it recovered completely. Immediately after recovery, fish were released back into the river at the place of capture. All the capture and handling procedures of fish were followed using standard ethical guidelines of North West University, South Africa (O'Brien et al. 2012a, 2013a, b).

We monitored tagged individuals using various techniques including foot walks along the banks of the Vaal River, small inflatable boats, and by air in a fixed wing aircraft (O'Brien et al. 2012a; 2013a, b; Fig. 2) from July 2007 to August 2010. We used only behavioural data collected after two weeks of release, although tagged largemouth yellowfish were monitored immediately to make sure there was survival and recovery from the tagging procedures (Bridger and Booth 2003; Rogers and White 2007). We carried out random and dedicated continuous 24-h surveys and the global positions of the tagged largemouth yellowfish (± 1 m accuracy) were recorded using a detailed georeferenced map of the site/hand-held global positioning system (GPS, eTrex (Garmin, Kansas City, USA), or GeoExplorer® 3000 Trimble (Trimble Navigation Ltd., Sunnyvale, CA, USA). Surveys were conducted for 3–4 days per month in a specific section of the river to cover the entire

study area for a year from the date of release. Individual fish were identifiable based on their unique radio transmitter signals visible on our computer/mobile system. We measured the total maximum movement distance of a tagged largemouth yellowfish during four consecutive 10-min intervals after locating the tagged fish. The movement categories were described as low (<10 m/10 min) and high movement (≥ 10 m/10 min). We evaluated activity: (feeding/non feeding), fish habitat cover (emergent and overhanging marginal vegetation, aquatic vegetation, boulders/rocks (merged boulders/rocks) and undercut banks (roots and dead/submerged trees), biotope (backwater areas, pools, glides, and rapids (riffling/runs), water column depth (mm) measured with a depth stick, water clarity (m) measured with a clarity tube, substrate types (silt, sand, gravel, cobble, boulder, bedrock), water colour, water surface flow (no flow, barely perceptible, riffle surface, smooth turbulent, undular broken) and associated it with the movement of yellowfish (Bovee 1986; Hirschowitz et al. 2007; King et al. 2010; O'Brien et al. 2012b; 2013a, b). Water flow (cumecs m^3/s), dissolved salts (mg/l) (Ca, Cl, K, Mg, and SO_4), dissolved nutrients (mg/l), (NO_3 , PO_4), dissolved toxicant (mg/l) (F, NH_4), pH, alkalinity (mg/l), electrical conductivity (mS/m), and Si (mg/l) were downloaded from the Department of Water and Sanitation, South Africa Affairs for the respective study areas.

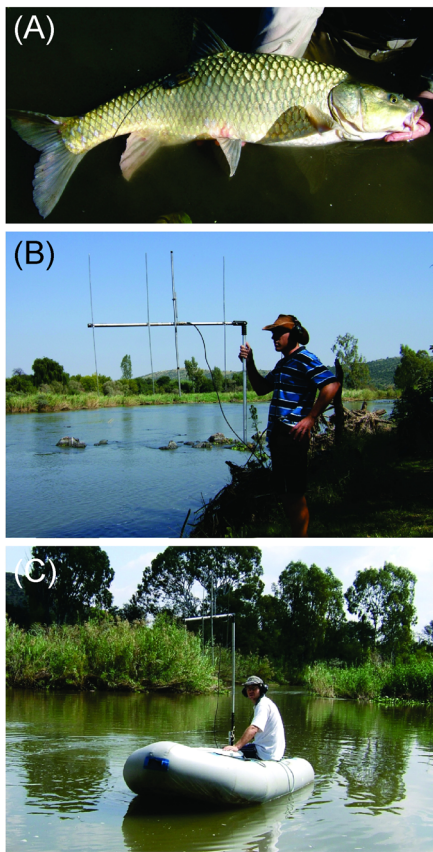


Fig. 2 Identifying the position of tagged largemouth yellowfish *Labeobarbus kimberleyensis* (a) by walking on the bank (b) and drifting in a boat (c) on the Vaal River, southern Africa

Data analyses

The movement was categorized as 0 or 1 if the movement was low (<10 m/10 min) or high (≥ 10 m/10 min) respectively. Factors associated with individual fish movement were identified using Binomial Generalised Linear Models (GLM) with mixed effects regression as a function of covariates using the lme4 package (Bates et al. 2014) in R (R Development Core Team 2014). Covariates and individual fish were used as fixed and random effects in the model, respectively. Habitat features and water quality were tested separately for their respective influence on largemouth yellowfish movement. To avoid issues with multicollinearity among predictor variables, we removed the correlated variables ($r > 0.60$) in a hierarchical approach using Pearson correlation co-efficient test (Graham 2003). First, we performed correlation tests among variables within

habitat features, and water quality variables separately then retained the least correlated habitat features (activity, cover, biotope, depth, water clarity, substrate and water surface flow) and water quality variables (Cl, F, NH₄, PO₄, pH, alkalinity and Si) for further statistical analysis.

The best-fit candidate models with few predictors were selected based on the framework of Burnham and Anderson (2002). Depth, water clarity, Cl, alkalinity and Si were log transformed (\log_{10}) prior to regression, allowing model coefficients to be interpreted as the change in the log-odds ratio (Cooch and White 2005). Model fit was assessed by examining Akaike's information criterion, standardized residuals, and observed vs. predicted values (Burnham and Anderson 1998). Significance of the individual regression coefficients was evaluated at $\alpha = 0.05$ level. Models were chosen based on p -values (significant when $p < 0.05$) of covariates, by examining plots of residuals and using Akaike's information criterion (AIC) to compare candidate models. The best model (the model with the minimum AIC value) was used as relative measure of model rank; models with delta AIC values less than 2 suggest substantial evidence for the model (Burnham and Anderson 2002). All candidate models ranking less than 2 delta AIC were used as a guide to select the best-fit models explaining movement of largemouth yellowfish (Burnham and Anderson 2002). Estimates of the relative importance of predictor variables were made by summing the Akaike weights (w_i) across all the models in which the variable occurred (Burnham and Anderson 2002). We evaluated the direction and effect of variables based on the average estimates of the parameter coefficient and its precision across the entire set of models (Burnham and Anderson 2002). All statistical analyses were done in Programme R version 3.0 (R Development Core Team 2014) using other supporting packages MASS (Venables and Ripley 2002), rJava (Urbanek 2010), glmulti (Calcagno and de Mazancourt 2010) and MuMIn (Bartoń 2013).

Results

Out of 27 largemouth yellowfish fitted with transmitters, we successfully tracked 12 individuals. The adult individual's mean morphometric data were as follows: mean body mass 3616 ± 220 g, mean total length $678 \pm$

14 mm, mean fork length 616 ± 12 mm and mean girth 370 ± 8 mm. The 12 tracked adult individuals resulted in 538 observations.

When we related the movement of 12 individuals to habitat features, the two best models were identified based on ≤ 2 delta AIC (Table 1). The relative variable importance across all the models were water clarity ($w_i = 1$), feeding ($w_i = 1$), habitat cover ($w_i = 0.96$) and biotope ($w_i = 0.35$) as main predictor variables for yellowfish movement, while the contribution of other variables was less ($w_i = \leq 0.01$). The coefficient estimate of the top two models was almost consistent and suggests that movement increased with decreasing water clarity ($\beta = -13.04 \pm 2.79$, $p < 0.005$), presence of feeding event ($\beta = 6.01 \pm 0.78$, $p < 0.005$), emergent and overhanging marginal vegetation cover ($\beta = 3.81 \pm 1.30$, $p = 0.003$) and rapid biotope ($\beta = 4.13 \pm 2.65$, $p = 0.120$). The movement reduced at water column but not significantly ($\beta = -2.33 \pm 1.36$, $p = 0.087$) among all habitat covers. Overall GLM mixed model coefficients showed a significant relationship between probability of largemouth yellowfish movement with water clarity, habitat cover and feeding event (Fig. 3).

The relationship between largemouth yellowfish movement and water quality was explored separately to evaluate the specific contribution of each variable. Two best candidate models were identified for probability of largemouth yellowfish movement based on the model with low AICc (≤ 2 delta AIC). Chloride (Cl) and PO_4 representing salt and nutrient variables were identified as two important predictors of movement across

the two competing models (Table 1). However, other important variables such as Si and alkalinity were also found to influence probability of high movement across the top models. Movement increased with increasing levels of Cl ($\beta = 57.44 \pm 7.17$, $p < 0.005$) and Si ($\beta = 3.61 \pm 0.47$, $p < 0.005$) whereas movement decreased with high PO_4 levels ($\beta = -35.83 \pm 7.58$, $p < 0.005$) and increased with alkalinity levels ($\beta = -20.19 \pm 13.81$, $p = 0.144$). The relative variable contribution across all models for Cl, PO_4 nutrient and Si as predictor variables was higher ($w_i = 1.0$ for each) than alkalinity ($w_i = 0.41$) and F toxicant ($w_i = 0.32$), whereas the contribution of other variables was minor ($w_i = \leq 0.06$). GLM coefficients showed a significant relationship between probability of largemouth yellowfish movement with Cl, PO_4 nutrient, and Si (Fig. 4).

Discussion

Globally fish species living in freshwater habitats are among the most endangered taxa on Earth (Collen et al. 2014). We found that largemouth yellowfish movements increased significantly with decrease in water clarity, presence of emergent and overhanging marginal vegetation, rapid biotope and presence of feeding events. When water quality was considered, largemouth yellowfish movement increased with increasing levels of Cl and Si whereas their movement decreased with high PO_4 nutrient load and increased with alkalinity levels in the river. In particular riverine fish species

Table 1 The top generalised linear mixed effect multi models showing the effect of habitat features and water quality variables on movement response of large mouth yellowfish *Labeobarbus kimberleyensis*

Environmental stressors	Selected models	df	logLik	AICc	delta AIC	Weight
Habitat features	Water clarity + Feeding + Available habitat	8	-112.09	240.45	0.00	0.61
	Biotope+ Water clarity + Feeding + Available habitat	11	-109.52	241.55	1.09	0.35
	Water clarity + Feeding	4	-119.57	247.22	6.76	0.02
	Water clarity + Depth + Feeding	5	-119.47	249.05	8.60	0.01
	Water clarity + Depth + Feeding + Substrate	6	-119.04	250.23	9.77	0.00
Water quality (chemical variables)	Cl + PO_4 + Si	5	-195.1	400.32	0.00	0.39
	Cl + PO_4 + Si + Alkalinity	6	-194.34	400.84	0.52	0.30
	Cl + F + PO_4 + Si	6	-195.08	402.32	2.00	0.14
	Cl + F + PO_4 + Si + Alkalinity	7	-194.29	402.79	2.47	0.11
	Cl + F + pH + PO_4 + Si	7	-194.87	403.96	3.64	0.06

df, residual degrees of freedom; logLik, Log likelihood; AIC, Delta Akaike Information Criterion; Delta AIC, the difference in AIC values between each model and the model with the lowest AIC; PO_4 , Dissolved Phosphorus; Cl, Chloride; Si, Silicon; F, Fluoride

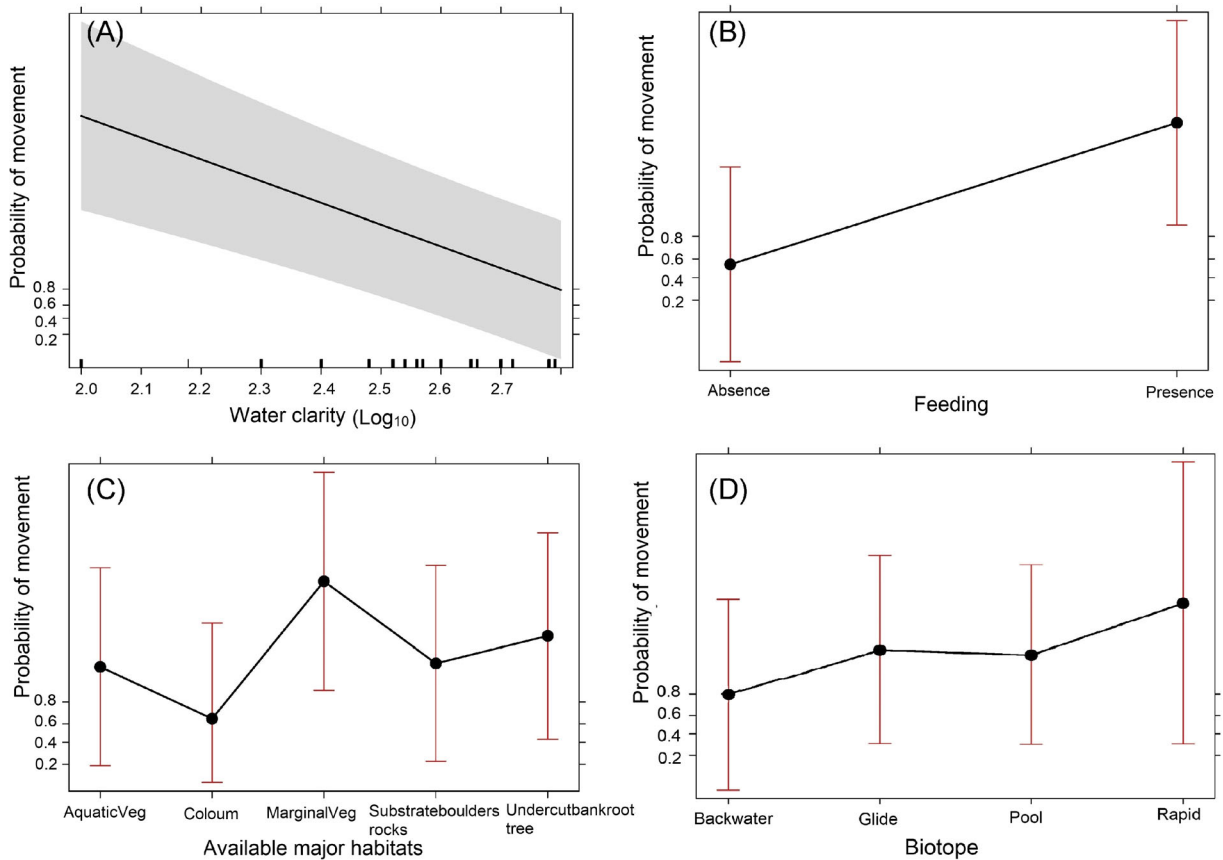


Fig. 3 Generalised linear mixed effect model (\pm 95% confidence intervals) explaining the predicted relationships between movement of large mouth yellowfish *Labeobarbus kimberleyensis* and

covariates (water clarity (a), occurrence of feeding event (b), available major habitat (c) and biotope (d) from the best models ($\leq 2\Delta AIC$)

faced with critical habitat degradation by a variety of anthropogenic disturbances, such as dam and weir construction and accumulation of heavy metals, nutrients, other chemicals from industries and urban waste, and runoff of pesticide use in the nearby agricultural land (Rahel et al. 1996; Bervoets and Blust 2003; Nilsson et al. 2005; Habit et al. 2007; Wepener et al. 2011; Cooke et al. 2012). The threatened riverine fish species such as largemouth yellowfish are more vulnerable to these impacts because of its sensitivity to degraded water quality (Impson and Swartz 2007). These detrimental effects affect fish metabolic activity, resistance to diseases, reproductive potential, and ultimately the health and survival of individual/population (Chapman 1996; Barton 2002).

Water clarity is one of the indicators of habitat condition of freshwater ecosystems (Geisler et al. 2016). Variations in water clarity influence surface temperature, heat budgets and stable environment in the freshwater

system (Mazumder and Taylor 1994; Fee et al. 1996). The amount of light passing through the water column is needed for photosynthesis (Walmsley and Bruwer 1980; Dallas and Day 2004). Furthermore, poor land use practices around Vaal River causes soil erosion that leads to elevated turbidity (Mulder 1973). Consequently, turbid environments affect primary production (Maitland 1995), zooplankton production (Walmsley and Bruwer 1980), and subsequently reduces the food availability and oxygen levels for fish (Kirk and Akhurst 1984; Newcombe and MacDonald 1991). Many fish species use visual cues to select feeding sites, capture prey and escape from potential threats (Aksnes and Utne 1997; Vogel and Beauchamp 1999; Figueiredo et al. 2015). The largemouth yellowfish as a visual predator, relies on water visibility while feeding. Particularly, such turbid environment significantly decreases feeding success rates of piscivorous fish, which feed on more visible prey (Mulder 1973; De Robertis et al.

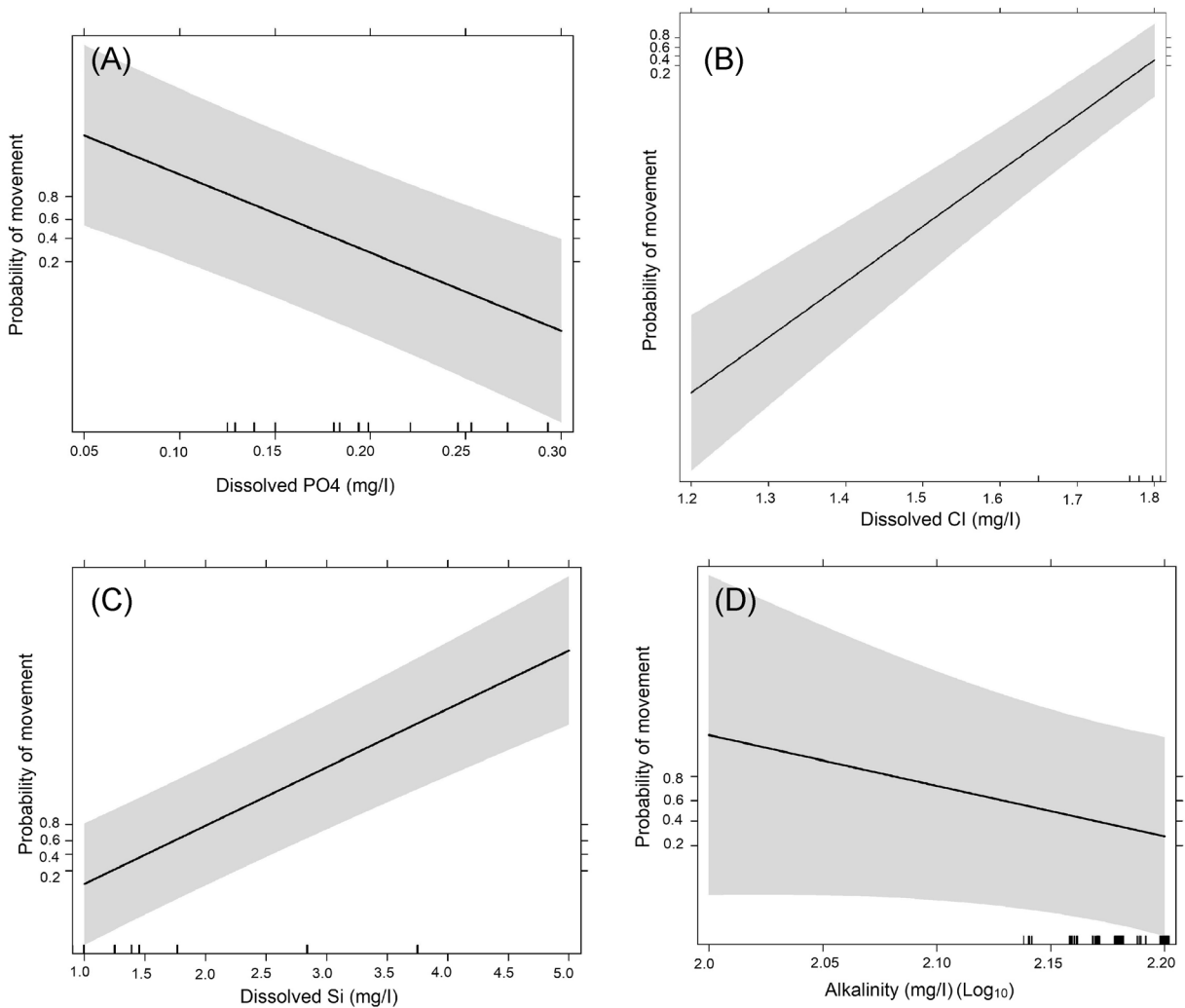


Fig. 4 Generalised linear mixed effect model (\pm 95% confidence intervals) explaining the predicted relationships between movement of largemouth yellowfish *Labeobarbus kimberleyensis* and

covariates (dissolved Phosphate (PO₄; **a**), Chloride (Cl; **b**), Silicon (Si; **c**) and Alkalinity (**d**) from the best models ($\leq 2\Delta AIC$)

2003; Figueiredo et al. 2015). Generally, low prey capture success rate increases the high mobility in fish (Gregory and Levings 1996; De Robertis et al. 2003). Therefore, the reduced feeding efficiency might have enhanced the high movement in largemouth yellowfish, and this may have an impact on the physiology of the species (Dallas and Day 2004).

We found that largemouth yellowfish movement was higher in vegetated areas. This could be due to higher availability of habitats which favours environmental quality (Gregg and Rose 1985; Schultz and Dibble 2012), emergent and overhanging marginal vegetation provides cover reducing mammal and bird predation (Uieda and Motta 2007) and can help yellowfish by

avoiding exposure to sunlight, wind action, temperature variation and water flow in the freshwater system (Boss and Richardson 2002). Our study revealed high movement of largemouth yellowfish in emergent and overhanging marginal vegetation in the littoral zones and on the river bank probably because structural habitat complexity influences prey abundance and diversity (O'Brien et al. 2013a) and therefore, enforcing fish activity.

Our study found that largemouth yellowfish showed high movements when associated with the rapid biotope. This species prefers fast-flowing waters with sandy or rocky substrate (Mulder 1973; Skelton and Bills 2007) and shows strong behavioural response

patterns to disrupted/sudden changes in flow (O'Brien et al. 2013a). These responses possibly involved the coordinated high movement of individuals into suitable refuge areas to avoid rapidly changing water levels/water flow and sudden surface-associated temperature changes (O'Brien et al. 2013a). Size-selective predators such as otter *Aonyx* spp. and predatory birds preferentially consume larger fish (Matthews 1998) and an increased predation risk in fast flowing shallower areas of a river might account for the high movements of large size-classes of largemouth yellowfish.

In our study, we found low movement of largemouth yellowfish in response to high dissolved phosphorus nutrient. This was probably due to high nutrient load in the Vaal River (De Villiers and Thiar 2007; Wepener et al. 2011). Although nutrients such as dissolved phosphorus (P) and nitrogen (N) are important components for healthy aquatic ecosystems, they are of most concern because they can stimulate/limit the growth of algae and aquatic weeds that can lead to eutrophication (Graham and Wilcox 2000; Anderson et al. 2002; Nhapi and Tirivarombo 2004; Peretyatko et al. 2007; Villiers and Thiar 2007; Wepener et al. 2011; Akbarimehr et al. 2016). Consequently, the water quality can be further reduced when bacteria consume dead algae and use available dissolved oxygen and elevate ammonium levels, which hamper aquatic life (Hellawell 1986). Such environmental changes affect pH and temperature (Nhapi and Tirivarombo 2004). Therefore, hypoxic conditions reduce fish metabolic rate and movement in order to minimize energy expenditures (Schurmann and Steffensen 1994; Dallas and Day 2004; Herbert and Steffensen 2005; Zhang et al. 2009).

Elevated levels of dissolved Cl observed in the Vaal River during this study that exceeded the Target Water Quality Guideline levels of aquatic ecosystems in South Africa (DWA 1996), increased the probability of largemouth yellowfish movement in the Vaal River during our study. Water salinity is an important natural stressor affecting aquatic communities in the freshwater ecosystem (Gutiérrez-Cánovas et al. 2009; Vidal-Abarca et al. 2013; Schriever et al. 2015). Dissolved Chloride (Cl) is one of the major ions found in most natural waters and its levels increase due to weathering of rocks, industrial and sewage effluents, and the use of chlorination processes for drinking where runoff occurs into rivers (Wannamaker and Rice 2000). Thus, these polluted aquatic systems can stimulate high movement

of largemouth yellowfish. Similarly, we also found significant increase in fish movement in response to increased levels of Si. Although, Si is removed from water naturally through reverse weathering process such as plankton fixation and sediment settling of dissolved silicon with clay minerals (Tallberg 2000), high concentrations of Si can also significantly influence and limit phytoplankton communities under eutrophication (Tallberg 2000; Conley et al. 1993). Alkalinity was not a significant predictor determining the largemouth yellowfish movement in the Vaal River.

Conservation implications

Largemouth yellowfish can serve as an appropriate indicator of the effect of these environmental variable changes as they exhibited complex behavioural responses to environmental changes in mainstream riverine habitats. The major threats to largemouth yellowfish in the Vaal River are associated with industrial/mining effluents, agricultural return flow, unfiltered effluent from municipal waste water treatment plants and solid waste in the storm water return flow (Impson et al. 2008) hampering the water quality (Wepener et al. 2011). Our study indicated that water clarity, emergent and overhanging marginal vegetation, rapid biotope, nutrient load and salinity clearly have a pervasive influence on fish movement, facilitating efforts to predict how fishes respond to environmental change. Influence of such factors could affect persistence and colonisation rates of such sensitive fish species in the long-term. Based on this, movement behaviour of this species could be important for management and conservation actions in this riverine system. The relationship between ecological factors and fish movement can be considered of relevance for future experimental and field research. These components are crucial for subsequent water quality monitoring and predicting changes in fish populations as a direct result of anthropogenic disturbances (Merciai et al. 2014). Consequence of pollutants discharged in the aquatic environment is likely to accumulate and cause potential risk not only to the fish species but also to piscivorous bird and mammal species including humans. Hence, it is essential to minimise the major sources of phosphorus from runoff from agricultural fertilizers, manure, and organic sewage wastes and industrial effluent into a river (Mallin et al. 1993; Anderson et al. 2002; Nhapi and Tirivarombo 2004).

Further studies should focus on the relationship between largemouth yellowfish feeding and their migratory behaviour in relation to dam and weir construction, characterise and evaluate spawning habitats and conditions, drought/flood refuge and emergent and overhanging marginal/aquatic vegetation, seasonal movements w.r.t different life histories of the study species, may have important implications for colonization and persistence of river fish populations (Matthews 1998; Albanese et al. 2004). Therefore, our study indicated freshwater fish species like the largemouth yellowfish can act as sensitive ecological indicators and illustrate some of the present habitat degradation pressures on freshwater ecosystems needing urgent attention.

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Author's contributions TR, GCO and CTD conceptualised the paper. GCO conducted the field work. TR analysed the data. TR wrote the manuscript; GCO and CTD provided editorial advice.

Compliance with ethical standards Our research and animal tagging process was approved by the North West Ethics Committee (NWU-00095-12-A4).

Conflict of interest The authors declare that they have no conflict of interest.

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